# The growth of *Vibrio vulnificus* and the habitat of infected patients in Kumamoto

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**SUMMARY** In Japan, Vibrio vulnificus (V. vulnificus) infection is very rare, and most infections have occurred in Kumamoto Prefecture (1), and especially around the Ariake and Yatsushiro seas. To investigate the relationship between the occurrence of V. vulnificus infection and environmental factors, including the salinity of seawater and the amount of rain in the Ariake and Yatsushiro seas, we measured the most probable number (MPN) of V. vulnificus in seawater and sea mud. In the Ariake Sea, we also observed the temperature and salinity of seawater at one site located on an estuary where the salinity is easily affected by river water and another site located offshore where seawater is little affected by river water. Furthermore, we investigated the MPN of V. vulnificus and observed the temperature and the salinity of seawater in 25 sites in the Ariake and Yatsushiro seas from July to August 2003 and 2004. In addition, we collected data on patients with V. vulnificus infections in Kumamoto from 1990 to 2006. The MPN of V. vulnificus differed by sampling site. More V. vulnificus were detected around the inland sea than the open sea, and the increase in V. vulnificus levels was affected by rainfall around inland sea areas with many rivers. V. vulnificus increases significantly in brackish water areas, and the salinity of seawater was as important as the seawater temperature. In other words, an area's topography and amount of rain are believed to be important factors for the occurrence of V. vulnificus infection. V. vulnificus infection has been regarded as an infection of hot districts. However, the salinity of seawater may be more important than temperature for the growth of V. vulnificus. Therefore, investigating these geographical and meteorological factors can help predict areas with a higher number of V. vulnificus infection outbreaks.

Key Words: Vibrio vulnificus, tidelands, salinity of seawater, amount of rain, geographical factors, meteorological factors

### Introduction

In Japan, there have been over 200 cases of *V. vulnificus* infection reported since 1978 (2). There are still many questions about the occurrence of *V. vulnificus* infections.

The current authors treated 43 V. vulnificus patients

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Received August 9, 2007 Accepted November 30, 2007 between 1990 and 2006 in Kumamoto Prefecture (Figure 1). In addition, there are several reports on *V. vulnificus* infections in Nagasaki (3) and Saga (4) prefectures but there are none on Kagoshima or Miyazaki prefectures, even though these prefectures neighbor Kumamoto and have higher average temperatures. This may be attributed to the fact that in these prefectures there are few areas with brackish water where *V. vulnificus* can grow.

#### **Materials and Methods**

Samples were prepared by using subsurface seawater

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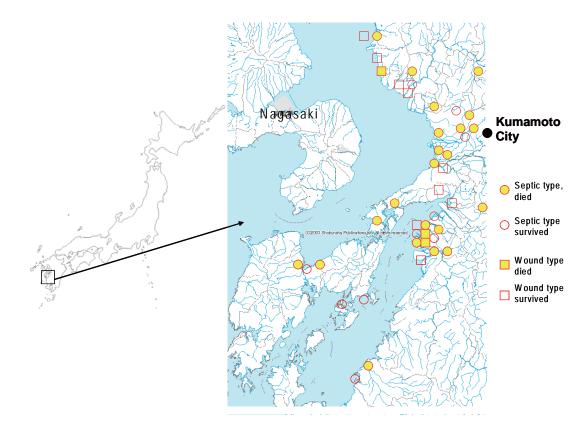


Figure 1. Patients with V. vulnificus infection in Kumamoto 1990-2006 (n = 43).

collected from Ariake Sea at a depth of between 1 m and 1.5 m. All samples were transported to the laboratory in insulated coolers and examined within 24-48 h of collection.

Seawater and mud samples were collected in sterilized glass bottles from sites A and B. Site A was located on the estuary of the Kikuchi River, where salinity is easily affected by river water, and site B, which was located offshore of the Ariake Sea 1.5 kilometers from Ooyano Island and where the salinity is little affected by river water. Samples were collected once a month from May to December, 2004.

Water samples were also collected in sterilized glass bottles from 25 different sites along the coast in Kumamoto (Figure 2) from July to August of 2003 and 2004.

The numbers of *V. vulnificus* in the seawater samples were estimated by the three-tube mostprobable-number (MPN) method. Volumes of 10 mL and 1 mL of seawater were added to 10 mL and 1 mL of double-strength alkaline peptone water (APW), respectively. One mL of dilutions  $(10^{-1}-10^{-4})$  in PBS was added to 10 mL APW. In addition, 500 mL of each seawater sample were filtered with a filter (pore size: 0.45 µm) and 40 mL APW were added to a tube containing the filter. After incubation at 35°C for 18 h, 10 µL of the culture were streaked onto CHROM agar Vibrio and incubated at 35°C for 18 h. Colonies

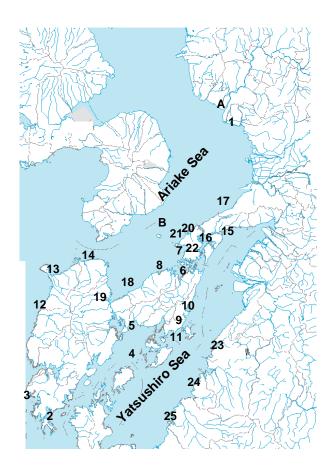


Figure 2. Water sampling sites along the coast in Kumamoto.

suspected of being *V. vulnificus* were confirmed to be *V. vulnificus* by the oxidase test, culture in triple sugar iron agar medium (Nissui Pharmaceutical Co. Ltd, Tokyo, Japan) containing 2% NaCl, culture in lysine indol motility medium (Nissui Pharmaceutical Co. Ltd) containing 2% NaCl, culture in VP semi-solid medium (Nissui Pharmaceutical Co. Ltd) containing 2% NaCl, culture in VP semi-solid medium (Nissui Pharmaceutical Co. Ltd) containing 2% NaCl, culture in VP semi-solid medium (Nissui Pharmaceutical Co. Ltd) containing 2% NaCl, and growth in nutrient broth (Oxoid, Hampshire, UK) containing 0, 3, 8, and 10% NaCl. Furthermore, the suspected colonies were tested for the presence of the cytotoxin-haemolysin gene of *V. vulnificus* by PCR with the primer set used by Hill *et al.* (5).

To serve as a collection stomacher, 500 g of sea mud were placed in a pouch that as then sealed and immediately transported to the lab. Sea mud in the sample poucher was mixed by hand and every bag was handled uniformly. Twenty-five-gram samples were thinly scattered in dishes, dried for 2 h at 110°C, and weighed after the sample was left in a desiccator for 40 min; then, the quantity of interstitial water was calculated. An additional 25 g of sea mud were diluted  $10 \times$  with PBS. This solution was added to 10 mL APW and diluted  $10 \times$  with PBS. *V. vulnificus* was identified and the MPN of the interstitial water per 100 mL was calculated by the method described above.

The seawater samples were collected in sterilized plastic bottles. Salinity was measured with a salt analyser (SAT-210, Toa Electronics, Tokyo, Japan). The temperature of seawater was measured with a thermometer. Seawater salinity (19-31‰) was not constant and may have been affected by weather or other factors rather than seasonal factors, although the temperature seemed to be affected by seasonal factors.

#### Results

The MPN of *V. vulnificus* in seawater and sea mud from May to December in 2004 is shown in Table 1. *V. vulnificus* numbers started to increase in seawater in May when the seawater temperature exceeded 20°C

Table 1. V. vulnificus levels at Site A and Site B

Month	Site A				Site B			
	MPN (/100mL)		Sea water	Salinity of	MPN (/100mL)		Sea water	Salinity of
	Seawater	Sea mud*	Temperature (°C		Seawater	Sea mud*	Temperature (°C)	seawater (‰)
May	7	230,000	19	18.4	< 3	< 3	19.5	33.5
June	< 3	700	24.3	34.2	< 3	< 3	29	34.1
July	2,300	43,000	30.5	21.1	< 3	< 3	28	35.2
August	2,300	1,500	31	34.1	3	30	29.5	35.9
September	1,500	230,000	29.5	30.8	< 3	< 3	27	33.3
October	75	21,000	24	32.9	< 3	40	22.4	32.3
November	3	290	21	30	< 3	< 3	19	32.7
December	< 3	150	17.5	30.7	< 3	< 3	17	29

\*MPN of interstitial water (/100mL)

Table 2. Vibrio vulnificus at 25 sites around Kumamoto (from July to August in 2003 and 2004)

Site	2003			2004		
	MPN (/100mL)	Amount of rain (mm)*	Salinity of seawater (‰)	MPN (/100mL)	Amount of rain (mm)*	Salinity of seawater (‰)
1	23,000	180	19.6	2300	0	34.1
2	< 3	30	33.7	< 3	0	37.3
3	< 3	30	33.5	< 3	0	37.8
4	< 3	35	31.8	< 3	0	36.5
5	7	174	30	< 3	0	37.2
6	< 3	208	26.1	< 3	24	34.8
7	4	208	26.1	3	24	34.7
8	4	208	25.3	3	24	34.4
9	4	205	21.4	< 3	4	34.1
10	3,800	205	22.1	< 3	4	34.4
11	< 3	205	24	< 3	4	35
12	< 3	129	33	< 3	6	36
13	< 3	30	32.7	< 3	6	36.2
14	< 3	174	30.8	< 3	6	35.5
15	93	128	25.6	< 3	6	35.3
16	430	128	24.1	< 3	6	36.5
17	230	128	26.8	3	6	35.9
18	< 3	278	32.7	3	34	35.9
19	< 3	174	31.3	< 3	4	37
20	36	44	31.9	< 3	70	35.8
21	6	44	31.3	< 3	70	36.4
22	< 3	44	31.7	< 3	70	36.4
23	21	251	30.3	< 3	30	35.6
24	9	251	31.6	< 3	0	35.4
25	23	204	31.6	< 3	2	35

\* Amount of rain during the seven days before each seawater sampling

at site A, and the peak number of *V. vulnificus* was from July to October when the seawater temperature exceeded 30°C. *V. vulnificus* inhabits sea mud more so than seawater, *V. vulnificus* was only isolated from seawater once in August at site B. In addition, the MPN was low, as shown in Table 1. *V. vulnificus* was isolated from sea mud in August and October, and the numbers were higher than from seawater. In addition, the salinity was stable. The increase in *V. vulnificus* was probably affected by the salinity, and *V. vulnificus* may have increased in number in sea mud more so than in seawater.

The MPN of *V. vulnificus* at twenty-five sites in Kumamoto in July and August in 2003 and 2004 is shown in Figure 1 and Table 2. The amount of rain in Table 2 shows the total rain during the seven days before each seawater sampling. There was more rainfall in 2003 than in 2004. In 2003, *V. vulnificus* was isolated

from 14 of 25 sites but from only five sites in 2004. The salinity of seawater was higher in 2004 than in 2003 at all of the sites investigated. The difference in seawater salinity could have been caused by a difference in rainfall.

Kumamoto is one area where numerous *V. vulnificus* infections occur. Since 1990, 30 patients with *V. vulnificus* infection were seen at Kumamoto University Hospital and 13 patients were seen at 7 other hospitals in Kumamoto Prefecture (Table 3). However, there were no patients from islands facing the open sea.

## Discussion

*V. vulnificus* infection is very rare, but its prognosis is poor once it occurs. *V. vulnificus* infection is divided into several clinical types: septic, wound, digestive, and other types. In Asian countries, including Japan, many

 Table 3. V. vulnificus infection patients in Kumamoto from 1990 to 2006

No.	Year	Female/Male	Years	Туре	Day of onset	Died/Suvived
1	1990	F	58	Septic type	1990.10.30	D
2	1991	М	56	Septic type	1991.7.8	S
3	1992	М	48	Septic type	1992.8.6	D
4	1993	F	64	Septic type	1993.8.15	D
5	1995	М	44	Wound type	1995.7.12	D
6		М	57	Septic type	1995.7.13	D
7	1996	М	55	Septic type	1996.7.22	D
8		М	56	Ingestive type	1996.7.31	S
9		М	67	Septic type	1996.9.18	D
10	1997	М	35	Wound type	1997.6.13	D
11		М	53	Septic type	1997.7.21	D
12	1999	F	66	Septic type	1999.6.6	D
13		F	74	Wound type	1999.7.8	S
14		М	38	Septic type	1999.8	D
15		М	67	Septic type	1999.9.21	D
16	2001	М	61	Septic type	2001.6.29	D
17		М	72	Septic type	2001.7.10	S
18		М	61	Septic type	2001.7.10	S
19		М	77	Septic type	2001.7.4	S
20		М	62	Wound type	2001.7.7	S
21		М	56	Septic type	2001.7.12	D
22		М	43	Septic type	2001.7.17	D
23		М	74	Wound type	2001.7.18	S
24		М	77	Wound type	2001.8.	S
25		М	68	Septic type	2001.8.14	D
26	2002	М	67	Septic type	2002.8.23	D
27		М	70	Septic type	2002.9.19	S
28		М	63	Wound type	2002.10.6	S
29	2003	М	77	Wound type	2003.7.24	D
30		М	77	Wound type	2003.8.15	S
31		F	70	Septic type	2003.9.5	S
32		М	32	Wound type	2003.9.7	S
33		М	70	Septic type	2003.10.8	D
34	2004	М	58	Septic type	2004.6.17	D
35		М	40	Septic type	2004.6.21	S
36		М	66	Septic type	2004.9.21	S
37	2005	М	59	Septic type	2005.7.15	D
38		М	61	Septic type	2005.7.12	D
39		М	59	Wound type	2005.7.28	S
40		М	55	Septic type	2005.7.30	D
41		М	70	Wound type	2005.9.25	S
42	2006	М	69	Septic type	2006.7.19	D
43		М	70	Septic type	2006.8.3	D

patients have the septic type, which is assumed to be a result of eating fresh marine products. *V. vulnificus* infections have been reported all over the world (6-15) and especially in warm areas in the summer. The water temperature suitable for growth of *V. vulnificus* is more than 20°C (16), and adequate salinity is 15-25‰ (17), but its growth is suppressed when salinity exceeds 25‰. However, there are few studies of *V. vulnificus* in relation to environmental factors and therefore, many questions remain regarding the association of *V. vulnificus* and environmental factors.

There are few reports on *V. vulnificus* in sea mud. The current investigation found a higher concentration of *V. vulnificus* in sea mud than in seawater. For this reason, wound-type *V. vulnificus* infection may occur from sea mud without the patient even being in seawater. Furthermore, there was no clear difference between sites A and B in terms of the seawater temperature; however, the salinity changed greatly at site A in comparison to site B, and a much higher concentration of *V. vulnificus* was found at site A than site B. The salinity of seawater may be as important for the growth of *V. vulnificus* as seawater temperature.

Hoi et al. (18) isolated V. vulnificus from seawater in Denmark, and V. vulnificus grew rapidly with a higher seawater temperature. However, there were no changes in the salinity of seawater in Denmark. Hervio-Heath et al. (19) performed an environmental study in France and clearly confirmed the presence of V. vulnificus in Girond littoral in the Bordeaux district; salinities there were 10-20‰, which were lower than in other areas. However, the rate of detection of V. vulnificus is assumed to be low, and the salinity of Mediterranean seawater is more than 35‰, which is higher than in other seas. This could be the reason why there are few patients with V. vulnificus in areas around the Mediterranean Sea (20). V. vulnificus was detected in Chesapeake Bay (21) even at a time when the seawater temperature was less than 10°C. However, in North Carolina (22), it was detected only when seawater temperature was more than 20°C. The current study detected high levels of V. vulnificus in an inland sea even when seawater temperature was under 15°C (data not shown). Motes et al. (23) reported that the MPN of V. vulnificus is very high in the Arabian Gulf Coast region. the above reports suggest that the salinity of seawater is important for the growth of V. vulnificus.

The current authors reported the first outbreak of *V. vulnificus* infection in Japan in 2001 (24) and focused on rainfall and related factors at the time of the outbreak. Since 1990, forty-three patients have been confirmed to have the infection (Table 3) in Kumamoto. Patients were concentrated in a coastal area where there is a large river.

At sites 15, 16, and 17 in the peninsula area, the amount of rainfall in July to August 2003 was 128 mm and that in 2004 was 6 mm. The salinity in 2003

was between 24.1‰ and 26.8‰ and that in 2004 was between 35.3‰ and 36.5‰. The MPN of *V. vulnificus* in 2003 and 2004 was from 93 to 430 and <3, respectively. At sites 2, 3, 12, 13, and 14 in island areas, in contrast, the salinity of seawater was stable though there was much more rainfall in 2003 than in 2004, and *V. vulnificus* was not detected either in 2003 or 2004. Based on these results, *V. vulnificus* is unlikely to grow in island areas facing the open sea but instead proliferate in island seas following rain. In actuality, no patients with *V. vulnificus* were found around sites 2, 3, 12, 13, and 14 in island areas even though the inhabitants eat a great deal of raw fish.

Thus, the amount of rain and the salinity of seawater are believed to be associated with the occurrence of a *V. vulnificus* infection as much as seawater temperature. Geographical and meteorological factors are believed to greatly influence the occurrence of *V. vulnificus* infection.

The onset of this illness is abrupt, rapidly progressing to septic shock with a high mortality rate. Clinicians managing patients with chronic liver disease need to educate their patients about the risk associated with the consumption of raw shellfish.

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#### References

- Wickboldt LG, Sanders CV. *Vibrio vulnificus* infectioncase report and update since 1970. J Am Acad Dermatol 1983; 9:243-251.
- Hlady WG, Klontz KC. The epidemiology of Vibrio infections in Florida, 1981-1993. J Infect Dis 1996; 173:1176-1183.
- Kawano S, Matsuo T, Ikeda T, *et al.* Fulminating halophilic vibrio septicemia - an autopsy case. Saishinigaku 1978; 33:1243-1248. (in Japanese)
- Kojo Y, Johno M, Nakagawa K, *et al. Vibrio vulnificus* infection: strategy for diagnosis and treatment. Jpn J Dermatol 1999; 16:61-66.
- Hill WE, Keasler SP, Trucksess MW, Trucksess MW, Feng P, Kaysner CA, Lampel KA. Polymerase chain reaction identification of *Vibrio vulnificus* in artificially contaminated oysters. Appl Environ Microbiol 1991; 57:707-711.
- Klontz KC, Lieb S, Schreiber M, Janowski HT, Baldy LM, Gunn RA. Syndromes of Vibrio vulnificus infections-clinical and epidemiologic features in Florida cases, 1981-1987. Annals Internal Med 1988; 15: 318-323.
- Park SD, Shon HS, Joh NJ. *Vibrio vulnificus* septicemia in Korea: clinical and epidemiologic findings in seventy patients. J Am Acad Dermatol 1991; 24:397-403.
- 8. Collier DN. Cutaneous infections from coastal and marine bacteria. Dermatol Ther 2002; 15:1-9.

- Chuang YC, Yuan CY, Liu CY, Lan CK, Huang AH. Vibrio vulnificus infection in Taiwan: report of 28 cases and review of clinical manifestations and treatment. Clin Infect Dis 1992; 15:271-276.
- Hsueh PR, Lin CY, Tang HJ, Lee HC, Liu JW, Liu YC, Chuang YC. *Vibrio vulnificus* in Taiwan. Emerging Infect Dis 2004; 10:1363-1368.
- Chan TYK. Vibrio vulnificus infections in Asia: an overview. Southeast Asian J Trop Med Health 1995; 26:461-465.
- Joynt GM, Gomersall CD, Lyon DJ. Severe necrotizing fasciitis of the extremities caused by *Vibrionaceae*: experience of a Hong Kong tertiary hospital. Hong Kong Med J 1999; 5:63-68.
- 13. Maxwell EL, Maxwell BC, Pearson SR, Stanley PA. A case of *Vibrio vulnificus* septicaemia acquired in Victoria. Med J Australia 1991; 154:214-215.
- Dalsgaard A, Frimodt-Møller N, Bruun B, Høi L, Larsen JL. Clinical manifestations and molecular epidemiology of *Vibrio vulnificus* infections in Denmark. Eur J Clin Microbiol Infect Dis 1996; 15:227-232.
- Nair NV, Sengupta DN, Ghosh S. Halophilic vibrios from fish and meat in Calcutta. Indian J Med Res 1975; 63:558-564.
- 16. Kaspar CW, ML Tamplin. Effects of temperature and salinity on the survival of *Vibrio vulnificus* in seawater and shellfish. Apple Environ Microbiol 1993; 59:2425-2429.
- 17. Kelly MT. Effect of temperature and salinity on Vibrio

(Beneckea) *vulnificus* occurrence in a Gulf Coast environment. Appl Environ Microbiol 1982; 44:820-824.

- Hoi L, Larsen JL, Dalsgaard I, Dalsgaard A. Occurrence of *Vibrio vulnificus* biotypes in Danish marine environments. Appl Environ Microbiol 1998; 64:7-14.
- Hervio-Heath D, Colwell RR, Derrien A, Robert-Pillot A, Fournier JM, Pommepuy M. Occurrence of pathogenic *vibrios* in coastal areas of France. J Appl Microbiol 2002; 92:1123-1135.
- Maugeri TL, Carbone M, Vera MT, Gugliandolo C. Detection and differentiation of *Vibrio vulnificus* in seawater and plankton of a coastal zone of the Mediterranean Sea. Res Microbiol 2006; 157:194-200.
- Pfeffer CS, Hite MF, Oliver JD. Ecology of *Vibrio* vulnificus in estuarine waters of eastern North Carolina. Appl Environ Microbiol 2003; 698:3526-3531.
- 22. Wright AC, Hill RT, Johnson JA, Roghman MC, Colwell RR, Morris JG Jr. Distribution of *Vibrio vulnificus* in Chesapeake Bay. Appl Environ Microbiol 1996; 62:717-724.
- Motes ML, Depaola A, Cook DW, Veazey JE, Hunsucker JC, Garthright WE, Blodgett RJ, Chirtel SJ. Influence of water temperature and salinity on *Vibrio vulnificus* in Northern Gulf and Atlantic Coast Oysters (Crassostrea Virginica). Appl Environ Microbiol 1998; 64:1459-1465.
- 24. Inoue Y, Matsui T, Ono T. An outbreak of *Vibrio vulnificus* infection in Kumamoto, Japan, 2001. Arch Dermatol 2004; 140:888-889.