

Effect of dehydroepiandrosterone on atherosclerosis in postmenopausal women

Siwei Zhang^{1,2,3,§}, Jing Zhou^{1,2,3,§}, Lijuan Li^{1,2,3}, Xinyao Pan^{1,2,3}, Jing Lin^{1,2,3}, Chuyu Li^{1,2,3}, Wing Ting Leung^{1,2,3}, Ling Wang^{1,2,3,*}

¹ Laboratory for Reproductive Immunology, Hospital and Institute of Obstetrics and Gynecology, Shanghai Medical College, Fudan University, Shanghai, China;

² The Academy of Integrative Medicine of Fudan University, Shanghai, China;

³ Shanghai Key Laboratory of Female Reproductive Endocrine-related Diseases, Shanghai, China.

SUMMARY In China, cardiovascular disease (CVD) has surpassed malignant tumours to become the disease with the highest mortality rate, and atherosclerosis (AS) is an important pathological cause of CVD. Dehydroepiandrosterone (DHEA) is the most abundant steroid hormone in circulating human blood and is a precursor of estrogen and androgen. DHEA is converted into a series of sex hormones in local peripheral tissues where it acts physiologically. DHEA also acts therapeutically, thereby avoiding the adverse systemic reactions to sex hormones. DHEA inhibits AS, thus inhibiting the development of CVD, and it improves the prognosis for CVD. The incidence of CVD in postmenopausal women is substantially higher than that in premenopausal women, and that incidence is believed to be related to a decrease in ovarian function. The current review analyzes the mechanisms of postmenopausal women's susceptibility to AS. They tend to have dyslipidemia, and their vascular smooth muscle cells (VSMCs) proliferate and migrate more. In addition, oxidative stress and the inflammatory response of endothelial cells (ECs) are more serious in postmenopausal women. This review also discusses how DHEA combats AS by countering these mechanisms, which include regulating the blood lipid status, protecting ECs (including coping with oxidative stress and inflammatory reactions of the vascular endothelium, inhibiting apoptosis of ECs, and inducing NO production) and inhibiting the proliferation and migration of VSMCs. As a result, DHEA has great value in preventing AS and inhibiting its progression in postmenopausal women.

Keywords dehydroepiandrosterone, atherosclerosis, postmenopause, vascular smooth muscle cells, endothelial cells, blood lipid

1. Introduction

Cardiovascular disease (CVD) is a common disease that jeopardizes the health of postmenopausal women (1) and atherosclerosis (AS) is the most critical pathological cause of CVD (2). Premenopausal women rarely suffer from CVD. However, the incidence of CVD in postmenopausal women is 2-6 times higher than that in premenopausal women of the same age group (3), due to its close relationship to a postmenopausal estrogen deficiency (4) (Figure 1). A study (5) involving 879 women suggested that menopause was significantly associated with the risk of developing carotid plaques. Females with an earlier onset of menopause (< 45 years) had a significantly higher atherosclerotic plaque volume than those with an intermediate (45-52 years) or later onset of menopause (> 52 years), irrespective of other

cardiovascular risk factors (6). The mean carotid intima-media thickness (CIMT) of the common carotid artery in postmenopausal women was significantly thicker than that in premenopausal women, with a mean difference of 0.068 mm (7). A recent prospective cohort study (8) also found that an elevated or persistently high level of A β 1-40, an aging peptide, is related to the rate of progression of subclinical AS in postmenopausal women and negatively correlated with levels of DHEA-S. An increasing number of women are prescribed hormone replacement therapy (HRT) after menopause or ovarian resection to prevent and treat CVD, osteoporosis, Alzheimer's disease, and other related long-term postmenopausal complications (9-12).

Dehydroepiandrosterone (DHEA) is the precursor of estrogen and androgen and is thought to prevent the development of AS (13). Dehydroepiandrosterone

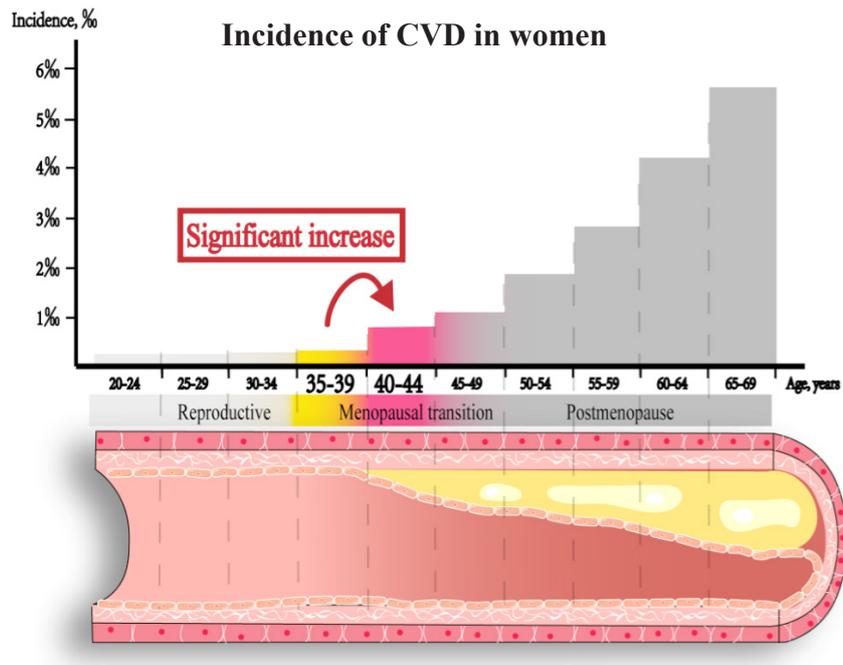


Figure 1. Incidence of CVD and vascular status in women. The incidence of CVD in women was stable at around 0.25%, and it increased significantly after the age of 40. The intima-media thickness of blood vessels in women also increased, with more lipid deposition.

sulfate (DHEA-S) is the metabolite of DHEA, and the level of DHEA-S is significantly inversely correlated with the incidence of CVD (14-17). For postmenopausal women with coronary risk factors, a lower DHEA-S level means a higher mortality due to CVD (18). The new immunosenescence paradigm proposed in recent years offers an explanation. Senescence leads to the loss of DHEA, which causes semi-activated macrophages to be immunosuppressed and unable to differentiate, while releasing pro-inflammatory cytokines in an unregulated manner. These dysfunctional cells accumulate in vascular tissue and lead to the development of AS (19). That said, DHEA also has positive effects on the brain, bones, emotions, and sexual function of postmenopausal women, so its clinical use warrants consideration.

2. Metabolism and pathway of DHEA

As early as 1934, DHEA was successfully separated from urine. In 1944, Munson discovered the sulfated form of DHEA (20). DHEA, also known as 3 β -hydroxyandrost-5-en-17-one, is the most abundant steroid circulating in human blood and is synthesized from cholesterol.

2.1. Generation of DHEA

The production of DHEA in the adrenal cortex and ovaries is regulated by adrenocorticotrophic hormone and gonadotropin, respectively. DHEA is mainly produced in the adrenal cortex, only 10% of DHEA is produced in the gonads, and the brain also produces a small amount of DHEA (21). Approximately 6-8 mg of DHEA are

produced per day in humans (22). In the blood, DHEA is mainly bound to albumin, a small amount will also bind to sex hormone-binding globulin (SHBG), and the remaining amount is free.

The level of DHEA changes during aging. The fetal adrenal gland produces a large amount of DHEA, but the level decreases rapidly after birth. The level of DHEA increases rapidly in the first two years of puberty, reaching a peak at 20-30 years of age, and then decreases at a rate of 2 to 5% annually. In individuals ages 70-80 years, the level of DHEA in the blood is only 10 to 20% of the peak level (23). The downstream hormones of the HPA axis have inhibitory feedback action on the upstream hormones, but DHEA does not participate in negative feedback regulation of the HPA axis. Thus, when the serum DHEA level is low, the body is unable to increase output through an endogenous feedback mechanism. Therefore, the body is unable to compensate for the deficiency in DHEA levels alone.

2.2. Conversion of DHEA

In the adrenal gland, endogenous DHEA is translated into DHEA-S by sulfation at the C3 β position. In addition, oral DHEA is converted into DHEA-S *via* the first pass effect of the liver and intestine. As mentioned above, DHEA-S is a circulating reservoir of DHEA. Circulating DHEA is transferred to related peripheral tissues (*e.g.*, the ovaries, prostate, bone, adipose tissue, and brain) and then converted into testosterone, androstenedione, estrone, dihydrotestosterone (DHT), and estradiol (E2).

DHEA has biological action locally and indirectly.

It is known to be a multi-directional "hormone buffer" and to supplement hormones in the body, which explains why it has been used to treat menopause-related diseases. At the same time, since only a small amount of DHEA is in free circulation and DHEA is only converted into estrogen in the peripheral tissues, systemic estrogen-like adverse effects, such as cholelithiasis (24) and venous thromboembolic and ischemic stroke events (25), can be avoided (26). In addition, when the level of DHEA in humans reaches 7 µg/L, the saturation of invertase occurs during the conversion of DHEA into active sex hormones, helping avoid a state of excess sex hormone levels in women.

2.3. Pathway of DHEA

DHEA and its oxidative metabolites have been found to activate some nuclear receptors, like the constitutive androgen receptor (AR), estrogen receptor (ER) alpha and beta, pregnane X receptor, peroxisome proliferator activated receptor (PPAR), and G protein-coupled ER (GPER1) (27,28). Since DHEA can be converted into androgen/estrogen, researchers have not clearly determined whether AR/ER are directly activated by DHEA or indirectly by the converted androgen/estrogen. However, the low affinity of ER for DHEA makes the latter a poor agonist of ER (27). In addition, the effects of DHEA on the proliferation and apoptosis of endothelial cells (ECs) and vascular smooth muscle cells (VSMCs) are not associated with ER and AR (29,30). DHEA and its analogues are not converted to estrogen or androgen but nonetheless have beneficial action on CVD, suggesting that DHEA interacts with its own specific receptor.

3. Pathogenesis of AS

Many theories on the mechanism of AS have been proposed from different perspectives. In recent years, most scholars have supported the "endothelial injury response" theory (31), which posits that endothelial dysfunction is an initial step in the pathogenesis of AS. The major risk factor for this disease is damage to the arterial intima, and the formation of atherosclerotic lesions results from the inflammatory-fibrotic proliferative response of arteries to intimal injury (Figure 2). AS is commonly regarded as a chronic inflammatory disease of the arterial wall caused by an imbalance in lipid metabolism and changes in inflammatory responses, whereby the body is unable to prevent the recruitment of inflammatory cells alone. However, a recent hypothesis suggests that AS is not just an inflammatory reaction of the blood vessel wall. Neither inflammatory cells nor necrotic cells are removed, and thus effector cell proliferation and tissue regeneration are eventually induced (32,33).

Blood vessels begin to change in the early stages of

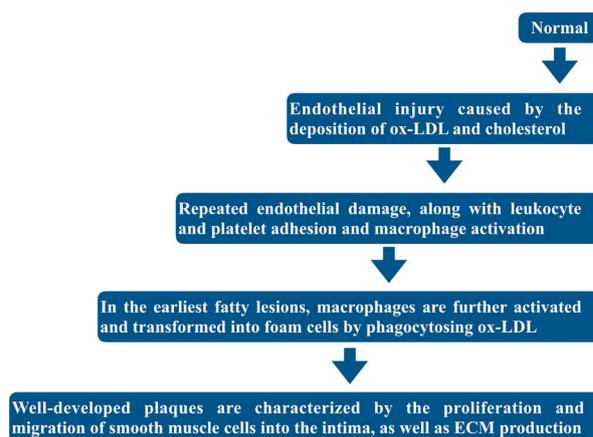


Figure 2. Pathogenesis of AS.

menopause. Endogenous estrogen and ERs decrease in postmenopausal women (34,35), resulting in the loss of inhibition of AS by estrogen, thus making them more vulnerable to AS. The mechanism may be an increase in the serum cholesterol level and high-density lipoprotein (HDL) particle size as well as interference with VSMC proliferation as a result of the decrease in endogenous estrogen and ERs (36).

4. The effects of DHEA on AS in postmenopausal women

4.1. DHEA alleviates dyslipidemia in postmenopausal women

Several early cross-sectional and prospective studies have revealed that the lipoprotein profile tends to worsen in postmenopausal women: plasma triglyceride (TG), total cholesterol (TC) low-density lipoprotein cholesterol (LDL-C), and lipoprotein levels increase and HDL cholesterol (HDL-C) levels decrease (37,38). In addition, studies (39,40) have indicated that age has more adverse effects on TC, LDL-C, TG, and non-HDL-C in postmenopausal women than BMI or smoking. This adverse change seems inevitable for postmenopausal women. However, dyslipidemia, which is mainly elevated LDL-C, is the most important factor for AS. Therefore, if AS in postmenopausal women is to be treated, then alleviating dyslipidemia is a very important aspect.

Substantial differences in the results of studies that have examined the effect of DHEA on blood lipid levels have been noted. Elevated plasma DHEA levels are reported to be correlated with HDL-C levels (41) but inversely correlated with LDL-C (42) and TC (43,44) levels. The correlation between plasma DHEA and TG levels was the most consistent. In a study by Jankowski *et al.*, treatment with DHEA resulted in a 17% reduction in serum TG levels (48). Lasco A *et al.* (42) reported that the serum TG levels of 20 postmenopausal women

decreased by about 20% after receiving DHEA (25 mg/d) for 12 months. A similar finding was noted in another study (45). However, one study (46) found that administration of DHEA does not change blood lipid parameters, which is consistent with the results of a previous study by the current authors (47). Use of lipid-lowering drugs may be a potential source of the inconsistency in the response of TG levels to DHEA (48). Therefore, whether DHEA can change blood lipid parameters or not needs to be studied more rigorously.

In addition to affecting blood lipid levels, DHEA can also directly inhibit lipid deposition (49). Fujioka *et al.* (50) found that DHEA can reduce the proliferation of adipocytes, which may be mediated by AR *via* an intracrine mechanism. DHEA also can promote lipid mobilization in adipose tissue by increasing the expression and activity of adipose triglyceride lipase (ATGL) and hormone-sensitive lipase (HSL) (51).

At present, there are conflicting results on improvement of blood lipid levels by DHEA. However, blood lipids are an important factor in the development and progression of AS, so the effect of DHEA on blood lipids needs to be studied further. Moreover, most of these studies involve normal people, and research needs to pay more attention to postmenopausal women.

4.2. DHEA corrects endothelial dysfunction in postmenopausal women

The loss of estradiol during postmenopausal may lead to a decline in endothelial function. For example, a decline in estradiol may alter the redox balance, thereby increasing oxidative stress and impairing endothelial function (52).

Endothelial dysfunction is involved in the pathogenesis of AS and CVD (53). One of the strategies for treating AS is to correct endothelial dysfunction (54). DHEA does not improve endothelial function through AR- or ER-mediated mechanisms (55,56). The effects of DHEA on ECs are shown in Figure 3.

4.2.1. DHEA inhibits EC oxidation

A study (57) has suggested that menopause is a risk factor for oxidative stress (OS). In postmenopausal women, not only progressive loss of estrogen and its protective effects (58), but also a further reduction in tocopherol and retinol levels as well as total antioxidant activity lead to OS (59). In a study by Taleb-Belkadi *et al.*, high levels of TBARS and carbonyl production and low levels of enzymatic defense found in

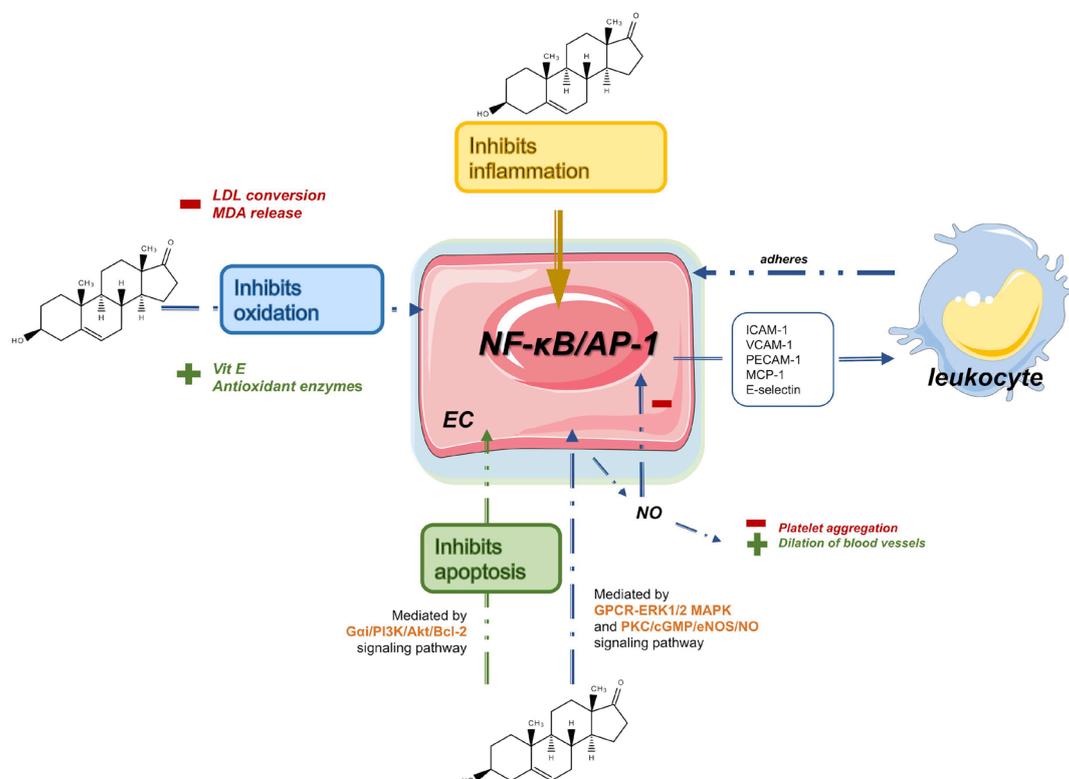


Figure 3. The effects of DHEA on ECs. First, DHEA inhibits EC oxidation by preventing the conversion of LDL to ox-LDL and the release of MDA, as well as by protecting endogenous vitamin E and the level and activity of antioxidant enzymes. Moreover, DHEA inhibits the production of MCP-1, ROS, ICAM-1, VCAM-1, PECAM-1, and E-selectin by ECs to prevent leukocytes from adhering to ECs, which involves NF-κB and AP-1. In addition, DHEA promotes NO production through the activation of eNOS *via* a GPCR-ERK1/2 MAPK cascade and the PKC/cGMP/eNOS/NO signalling pathway. NO subsequently inhibits platelet aggregation and the invasion and adhesion of leukocytes and it promotes the dilation of blood vessels. Moreover, DHEA protects ECs from apoptosis by activating the DHEAR/Gai/PI3K/Akt/Bcl-2 signalling pathway.

postmenopausal women indicated that the women were exposed to OS. OS, and especially the oxidation of LDL in the arterial wall, can lead to worse AS through the stages of the menopausal transition in healthy women (60). In addition, the production of the superoxide anion $O_2^{\cdot-}$ and an increase in the levels of peroxynitrite are also characteristics of atherosclerotic lesions (61,62).

According to a previous study (63), the synthesis of reactive oxygen species (ROS) promotes AS by increasing superoxide production and suppressing EC function. The production of large amounts of ROS overwhelms the antioxidant defenses in cells, causing neutrophil activation, protein modification, lipid peroxidation, and DNA damage, which are key factors that promote the development of AS and CVD (64,65) (Figure 3).

DHEA effectively inhibits the oxidation of low-density lipoprotein (LDL) to oxidized low-density lipoprotein (ox-LDL) (47,66), it inhibits ox-LDL-induced ROS production (67), it reduces superoxide production, it ameliorates endothelial dysfunction, and it prevents the development of AS.

In some experiments, DHEA increased the antioxidant capacity of LDL by protecting endogenous vitamin E (68) and by significantly reducing the chemotactic activity of monocytes (69), directly removing the free radicals produced by the lipoprotein oxidation process (70), and counteracting the cellular damage caused by LDL and ox-LDL, all of which enable DHEA to function as an antioxidant (66,68). In addition, DHEA restores the levels and activities of glutathione peroxidase, SOD, and catalase (71,72). Moreover, DHEA significantly inhibits the secretion of malondialdehyde (MDA) in ECs (47). As a cytotoxic end product of lipid peroxidation, MDA causes cross-linking polymerization of macromolecules such as proteins and nucleic acids and it affects the respiratory function of the mitochondria *in vitro*. At the same time, DHEA also increases the antioxidant capacity of certain subcellular structures (73).

In summary, DHEA has antioxidant action by inhibiting the production of ox-LDL and MDA, removing free radicals, reducing monocyte adhesion, and protecting antioxidant enzymes.

4.2.2. DHEA inhibits EC inflammation

The level of inflammation is higher in postmenopausal women, which is evident in higher levels of TNF- α , IL-1 α , and CRP (74,75). Novella *et al.* suggested that this may be due to the change in estrogen-mediated regulation of female inflammatory biomarkers (76) which were identified as independent risk factors for CVD in postmenopausal women (77). DHEA can reduce inflammation, and especially in ECs, and ECs are closely related to AS.

DHEA alleviates inflammation of ECs independent of the ER α or ER β pathway. *In vitro*, DHEA

significantly inhibits monocyte chemoattractant protein-1 (MCP-1) secretion, ROS production, and expression of intracellular adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule 1 (VCAM-1), platelet and EC adhesion molecule 1 (PECAM-1), and E-selectin (78). Moreover, DHEA also reduces the expression of adhesion molecule receptors in the U937 monocyte-like cell line, which suppresses the adhesion of monocytes to injured ECs (47). In one study (79), DHEA significantly reduced the LPS-induced transcription of nuclear factor kappa B (NF- κ B). Moreover, DHEA impairs monocyte adhesion by suppressing the activity of NF- κ B, thereby inhibiting the development of AS (47). A recent study (80) also indicated that DHEA restrains neutrophil recruitment and adhesion to ECs by reversing inflammation-induced down-regulation of developmental endothelial locus 1 (a secreted homeostasis factor) expression.

4.2.3. DHEA protects ECs by inducing NO production

The ability of vascular ECs to resist AS and antithrombotic factors largely relies on the production and release of active substances such as NO. NO blocks the expression of pro-inflammatory molecules as well as adhesion molecules in ECs. NO also inhibits the infiltration and adhesion of leukocytes (81).

Healthy endothelium, which normally produces NO, avoids the development and complications of AS (82). Nevertheless, the production of estrogen is reduced in postmenopausal women, and thus the activity of NO synthase decreases (83,84), which leads to a decrease in NO synthesis in ECs. A study found that a lack of NO and damaged endothelial progenitor cells resulted in vasodilation dysfunction in postmenopausal women, who are more prone to CVD, and especially AS.

DHEA activates eNOS through genomic and non-genomic mechanisms, and DHEA directly regulates human vascular walls by controlling the synthesis and stability of the eNOS protein in ECs (85). DHEA also effectively increases serum NO levels by activating PKC/cGMP/eNOS/NO pathways to prevent platelet aggregation, improve EC function, and alleviate early pathological changes associated with AS (44,47,86).

4.2.4. DHEA promotes EC proliferation and inhibits EC apoptosis

During aging, EC apoptosis increases, which affects the development of AS (87,88). The production of TNF- α induced by LPS and testosterone promotes apoptosis of ECs, whereas DHEA has the opposite effect on ECs. DHEA increases EC proliferation *in vitro* (44) and protects ECs from apoptosis (89). This anti-apoptotic effect of DHEA does not rely on ER or conversion into E2, but it is associated with the GTP-binding protein (G α i) and the downstream phosphatidylinositol 3-kinase

(PI3K)/Akt signalling cascade (90).

4.3. DHEA inhibits the proliferation and migration of VSMCs

Lee *et al.* (91) noted marked proliferation of aortic VSMCs in ovariectomized mice. During aging, the level of sirtuin 1, a novel modulator of neointima formation caused by arterial injury, decreased (92). The reduction in this protein indirectly promotes the proliferation and migration of VSMCs (93). VSMC proliferation and migration of surrounding extracellular matrix (ECM) are the main reasons for thickening of the intimal wall, which will lead to AS (94).

DHEA is involved in relaxing VSMCs and inhibiting the proliferation and migration of VSMCs (30,95). DHEA does not have a significant effect on the phenotypic transition of VSMCs but rather reduces OS and inflammation in VSMCs by directly interrupting the ROS-dependent ERK1/2 signalling and p38 mitogen-activated protein kinase (MAPK)/NF- κ B signalling pathways, thereby inhibiting the proliferation of VSMCs (95). Regardless of whether VSMCs undergo a phenotypic shift, DHEA can have a beneficial effect on these cells. DHEA-specific receptors are present in human VSMCs, and DHEA regulates the proliferation and apoptosis of VSMCs *via* a mechanism independent of ER and AR (30,44).

All of the aforementioned effects of DHEA on AS are shown in Figures 4 and 5 and Table 1.

5. Use of DHEA in the treatment of AS

As early as 1996, one study (102) proposed that DHEA is the source of youth, but the clinical use of DHEA is still hotly debated.

Several of the aforementioned studies have indicated that DHEA has anti-atherosclerotic action in animal models. DHEA improves cardiovascular risk-related parameters (42) and can be used as a drug for primary prevention of CVD (103). However, some studies have indicated that DHEA has no effect on CVD risk (104-106) and no effect on endothelial function (92,107-109). A meta-analysis (110) by Wu *et al.* noted no correlation between the level of DHEA-S and AS. However, other meta-analyses (15,111) noted that the lower the level of DHEA, the worse the prognosis for patients with CVD.

Qin *et al.* (38) suggested that DHEA had no effect on the blood lipid profile, and especially that of healthy postmenopausal women (112). This finding is consistent with the results of a previous study by the current authors (47). Nevertheless, there may be health benefits for women with adrenal insufficiency (113,114).

There are many factors responsible for the differing results of those studies. At present, many studies are based on rats and other rodents as models, but they are not the best model because they have almost no endogenous DHEA (115). In addition, the dosage of DHEA in those experiments is usually too high and it differs (112,113). Moreover, DHEA is rapidly metabolized, leading to somewhat differing results of many studies (43). A recent meta-analysis (116) suggested that publication bias and small flawed studies may also explain the discrepancy.

Therefore, whether postmenopausal women should

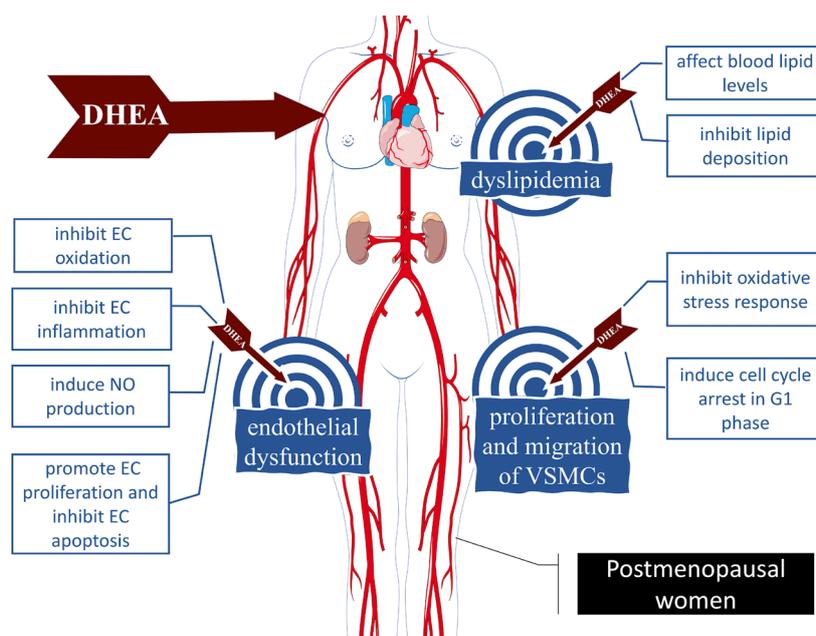


Figure 4. DHEA has specific action against aspects of AS in postmenopausal women. Postmenopausal women have dyslipidemia, abnormal proliferation and migration of VSMCs, and endothelial dysfunction. DHEA can play a role in alleviating these adverse aspects. First, it can improve dyslipidemia by affecting blood lipid levels and inhibiting lipid deposition. Second, DHEA can inhibit the oxidative stress response and induce cell cycle arrest in the G1 phase. In addition, DHEA can inhibit the oxidation and inflammation of ECs, induce NO protection, promote the proliferation of ECs, and inhibit the apoptosis of ECs.

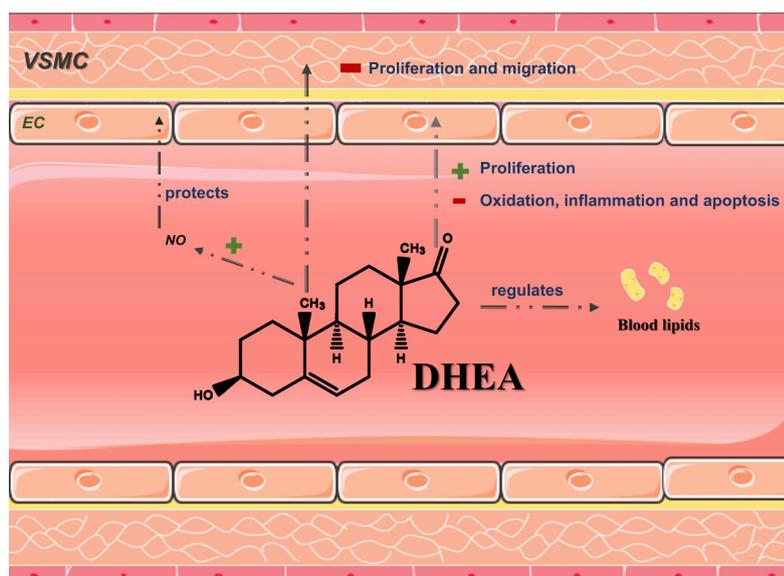


Figure 5. The effects of DHEA on atherosclerosis. First, DHEA affects the development and progression of AS by regulating blood lipid parameters, but substantial differences in results have been noted. Moreover, DHEA preserves EC function by inhibiting the oxidation and inflammation of ECs through NO production and promoting EC proliferation and inhibiting EC apoptosis. In addition, DHEA inhibits the progression of AS by inhibiting the proliferation and migration of VSMCs.

Table 1. The effects of DHEA on AS

Pathophysiological role of DHEA	Specific changes/mechanisms
Effects on blood lipids	A subject of debate
Effects on endothelial function	
Inhibition of EC oxidation	Prevention of LDL conversion to ox-LDL (47,66,67) Protection of endogenous vitamin E (68) Inhibition of leukocyte adhesion to ECs (69) Restoring the level and activity of antioxidant enzymes (70,72) Inhibition of MDA release by ECs (47,70)
Inhibition of EC inflammation	Inhibition of leukocyte adhesion to ECs: inhibiting the production of MCP-1, ROS, ICAM-1, VCAM-1, PECAM-1, and E-selectin by ECs; decreasing the expression of CCR2, LFA-1, and VLA-4 in the U937 monocyte-like cell line (47) Inhibition of IL-8, ICAM-1 and VCAM-1 production induced by TNF- α by blocking the LPS/TNF- α /PPAR α /NF- κ B signalling pathway (47,96) Inhibiting EC adhesion and oxidative stress by blocking AP-1 activity (67,97,98)
Protecting ECs through NO production	Inhibitory effect of NO on platelet aggregation and dilation of blood vessels (86) Inhibitory effect of NO on the expression of NF- κ B, ICAM-1 and VCAM-1; prevention of the invasion and adhesion of leukocytes (99) Activation of eNOS via a GPCR-ERK1/2 MAPK cascade (85) Increasing NO production through the PKC/cGMP/eNOS/NO signalling pathway (44,86)
Promotion of EC proliferation and inhibition of EC apoptosis	EC proliferation (85) Protecting ECs from apoptosis by activating the DHEAR/Gai/PI3K/Akt/Bcl-2 signalling pathway (89)
Inhibition of VSMC proliferation and migration	Promoting relaxation and inhibiting the proliferation of VSMCs by directly interrupting ROS-dependent ERK1/2 signalling and the p38 MAPK/NF- κ B signalling pathway (30,95) Inhibiting the phenotypic transition and proliferation of VSMCs by blocking platelet-derived growth factor receptor- β (PDGFR- β) and regulating glutathione/glutathione (GSH/GRX) and low molecular weight protein tyrosine phosphatase (LMW-PTP) (100) Causing apoptosis: inducing cell cycle arrest in the G1 phase; upregulating the expression of the cyclin-dependent kinase (CDK) inhibitor p16 ^{INK4a} , activating caspase-3, and inducing PPAR α expression in VSMCs (101)

take DHEA to treat or prevent forms of CVD such as AS is unclear. In addition, there is no clear standard for its indications and dosage (117).

DHEA causes adverse reactions such as hirsutism and acne. DHEA is believed to increase the risk of breast cancer in postmenopausal women (118,119). That said, experiments have indicated that the use of DHEA for 52

weeks has no effect on the endometrium (94). Evaluating the appropriate dose for patients is difficult because of the possibility of those adverse reactions, and indications for DHEA need to be carefully evaluated (120). Timing of use is also important. Treatment should start during menopausal transition, that is, within six years after menopause (93,121).

At present, studies on the clinical use of DHEA are still lacking. Therefore, use of DHEA should be carefully considered, the patient's eligibility should be determined, the patient's adrenal function should be considered, and whether the patient can tolerate the drug's adverse effects should be considered.

6. Conclusion

As a hormone precursor, DHEA is an endogenous steroid hormone and important source of estrogen and androgen in postmenopausal women. In addition, DHEA itself has a variety of biological actions that are independent of ER/AR and its conversion into estrogen/androgen, and it functions in almost all systems of the body (43). The current review has analyzed the mechanisms of postmenopausal women's susceptibility to AS. It has also discussed how DHEA plays a role in combating AS by countering these mechanisms, which include regulating the blood lipid status, protecting ECs (including coping with OS and inflammatory reactions of the vascular endothelium, inhibiting apoptosis of ECs, and inducing NO production) and inhibiting the proliferation and migration of VSMCs. In addition to its activity against AS, DHEA might have other protective effects on the cardiovascular system, such as preventing and reversing pulmonary hypertension (55) and reducing insulin resistance (122). However, further studies need to examine the mechanism and long-term effects of DHEA and additional clinical trials need to examine DHEA supplements. DHEA may serve as a better treatment for postmenopausal women and the entire population in the near future.

Acknowledgements

The authors sincerely appreciate the assistance of Peng Li and Suna Tian in preparing the figures.

Funding: This work was supported by grants from the 2018 Program to Guide Medicine ("Yixue Yindao") of the Shanghai Municipal Science and Technology Commission (grant no.18401902200 to L Wang), a project under the Scientific and Technological Innovation Action Plan of the Shanghai Natural Science Fund (grant no. 20ZR1409100 to L Wang), a project of the Chinese Association of Integration of Traditional and Western Medicine special foundation for Obstetrics and Gynecology-PuZheng Pharmaceutical Foundation (grant no. FCK-PZ-08 to L Wang), a project for hospital management of the Shanghai Hospital Association (grant no. X2021046 to L Wang), and a clinical trial project (L Wang) of the Special Foundation for Healthcare Research of the Shanghai Municipal Health Commission.

Conflict of Interest: The authors have no conflicts of interest to disclose.

References

- Pietrzak A, Czuczwar P, Mosiewicz J, Paszkowski T, Chodorowska G, Bartosinska J, Gerkowicz A, Paluszkiwicz P, Freud T, Cohen AD. Cardiovascular disease in psoriatic post-menopausal women. *J Eur Acad Dermatol Venereol.* 2015; 29:1231-1234.
- Pant S, Deshmukh A, GuruMurthy GS, Pothineni NV, Watts TE, Romeo F, Mehta JL. Inflammation and atherosclerosis-revisited. *J Cardiovasc Pharm T.* 2014; 19:170-178.
- Kannel WB, Hjortland MC, McNamara PM, Gordon T. Menopause and risk of cardiovascular disease: The Framingham study. *Ann Intern Med.* 1976; 85:447-452.
- Tuomikoski P, Mikkola TS. Postmenopausal hormone therapy and coronary heart disease in early postmenopausal women. *Ann Med.* 2014; 46:1-7.
- Li Y, Zhao D, Wang M, Sun JY, Liu J, Qi Y, Hao YC, Deng QJ, Liu J, Liu J, Liu M. Association of menopause with risk of carotid artery atherosclerosis. *Maturitas.* 2021; 143:171-177.
- Schreinlechner M, Noflatscher M, Reinstadler SJ, Sommer P, Lener D, Reiser E, Theurl M, Kirchmair R, Bauer A, Marschang P. Early onset of menopause is associated with increased peripheral atherosclerotic plaque volume and progression. *Atherosclerosis.* 2020; 297:25-31.
- Ieamtairat P, Soontrapa S, Kaewrudee S, Promsorn J, Takong W, Somboonporn W. Difference in carotid intima-media thickness between pre and postmenopausal women. *Menopause.* 2019; 26:39-44.
- Lambrinouadaki I, Delialis D, Georgiopoulos G, Tual-Chalot S, Vlachogiannis NI, Patras R, Aivalioti E, Armeni E, Augoulea A, Tsoltos N, Soureti A, Stellos K, Stamatelopoulos K. Circulating amyloid beta 1-40 is associated with increased rate of progression of atherosclerosis in menopause: A prospective cohort study. *Thromb Haemost.* 2021; 121:650-658.
- McCarrey AC, Resnick SM. Postmenopausal hormone therapy and cognition. *Horm Behav.* 2015; 74:167-172.
- Jackson RD, Mysiw WJ. Insights into the epidemiology of postmenopausal osteoporosis: The Women's Health Initiative. *Semin Reprod Med.* 2014; 32:454-462.
- Zhang J, Qiu X, Gui Y, Xu Y, Li D, Wang L. Dehydroepiandrosterone improves the ovarian reserve of women with diminished ovarian reserve and is a potential regulator of the immune response in the ovaries. *Biosci Trends.* 2015; 9:350-359.
- Lin J, Zhu J, Wang Y, Zhang N, Gober HJ, Qiu X, Li D, Wang L. Chinese single herbs and active ingredients for postmenopausal osteoporosis: From preclinical evidence to action mechanism. *Biosci Trends.* 2017; 11:496-506.
- Lee MJ, Kim EH, Lee SA, Kang YM, Jung CH, Yoon HK, Seol SM, Lee YL, Lee WJ, Park JY. Dehydroepiandrosterone prevents linoleic acid-induced endothelial cell senescence by increasing autophagy. *Metabolism.* 2015; 64:1134-1145.
- Hirokawa K, Ohira T, Nagayoshi M, Kajiura M, Imano H, Kitamura A, Kiyama M, Okada T, Iso H. Dehydroepiandrosterone-sulfate is associated with cardiovascular reactivity to stress in women. *Psychoneuroendocrinology.* 2016; 69:116-122.
- Mannic T, Viguie J, Rossier MF. *In vivo* and *in vitro* evidences of dehydroepiandrosterone protective role on the cardiovascular system. *Int J Endocrinol Metab.* 2015;

- 13:e24660.
16. Zhao D, Guallar E, Ballantyne CM, Post WS, Ouyang P, Vaidya D, Jia X, Ying W, Subramanya V, Ndumele CE, Hoogeveen RC, Michos ED. Sex hormones and incident heart failure in men and postmenopausal women: The atherosclerosis risk in communities study. *J Clin Endocrinol Metab.* 2020; 105:e3798-3807.
 17. Aribas E, Ahmadizar F, Mutlu U, Ikram MK, Bos D, Laven JSE, Klaver CCW, Ikram MA, Roeters van Lennep JL, Kavousi M. Sex steroids and markers of micro- and macrovascular damage among women and men from the general population. *Eur J Prev Cardiol.* 2021.
 18. Shufelt C, Bretsky P, Almeida CM, Johnson BD, Shaw LJ, Azziz R, Braunstein GD, Pepine CJ, Bittner V, Vido DA, Stanczyk FZ, Bairey Merz CN. DHEA-S levels and cardiovascular disease mortality in postmenopausal women: Results from the National Institutes of Health-National Heart, Lung, and Blood Institute (NHLBI)-sponsored Women's Ischemia Syndrome Evaluation (WISE). *J Clin Endocrinol Metab.* 2010; 95:4985-4992.
 19. Laderoute M. The paradigm of immunosenescence in atherosclerosis-cardiovascular disease (ASCVD). *Discov Med.* 2020; 29:41-51.
 20. Lieberman S. An abbreviated account of some aspects of the biochemistry of DHEA, 1934-1995. *Ann N Y Acad Sci.* 1995; 774:1-15.
 21. Ernst E. Textbook of natural medicine. Focus on Alternative and Complementary Therapies. 2010; 5:157-157.
 22. Couzinet B, Meduri G, Lecce MG, Young J, Brailly S, Loosfelt H, Milgrom E, Schaison G. The postmenopausal ovary is not a major androgen-producing gland. *J Clin Endocrinol Metab.* 2001; 86:5060-5066.
 23. Samaras N, Samaras D, Frangos E, Forster A, Philippe J. A review of age-related dehydroepiandrosterone decline and its association with well-known geriatric syndromes: Is treatment beneficial? *Rejuvenation Res.* 2013; 16:285-294.
 24. Wang S, Wang Y, Xu J, Chen Y. Is the oral contraceptive or hormone replacement therapy a risk factor for cholelithiasis: A systematic review and meta-analysis. *Medicine (Baltimore).* 2017; 96:e6556.
 25. Flores VA, Taylor HS. The effect of menopausal hormone therapies on breast cancer: Avoiding the risk. *Endocrinol Metab Clin North Am.* 2015; 44:587-602.
 26. Labrie F, Archer DF, Koltun W, *et al.* Efficacy of intravaginal dehydroepiandrosterone (DHEA) on moderate to severe dyspareunia and vaginal dryness, symptoms of vulvovaginal atrophy, and of the genitourinary syndrome of menopause. *Menopause.* 2016; 23:243-256.
 27. Clark BJ, Prough RA, Klinge CM. Mechanisms of action of dehydroepiandrosterone. *Vitam Horm.* 2018; 108:29-73.
 28. Klinge CM, Clark BJ, Prough RA. Dehydroepiandrosterone research: Past, current, and future. *Vitam Horm.* 2018; 108:1-28.
 29. Cai JJ, Wen J, Jiang WH, Lin J, Hong Y, Zhu YS. Androgen actions on endothelium functions and cardiovascular diseases. *J Geriatr Cardiol.* 2016; 13:183-196.
 30. Williams MR, Ling S, Dawood T, Hashimura K, Dai A, Li H, Liu JP, Funder JW, Sudhir K, Komesaroff PA. Dehydroepiandrosterone inhibits human vascular smooth muscle cell proliferation independent of ARs and ERs. *J Clin Endocrinol Metab.* 2002; 87:176-181.
 31. Yamada N. Atherosclerosis. *Nihon Rinsho.* 1999; 57:2345-2348.
 32. Viola J, Soehnlein O. Atherosclerosis - A matter of unresolved inflammation. *Semin Immunol.* 2015; 27:184-193.
 33. Gao B, Matsuura K, Shimizu T. Recent progress in induced pluripotent stem cell-derived cardiac cell sheets for tissue engineering. *Biosci Trends.* 2019; 13:292-298.
 34. El Khoudary SR, Thurston RC. Cardiovascular implications of the menopause transition: Endogenous sex hormones and vasomotor symptoms. *Obstet Gynecol Clin North Am.* 2018; 45:641-661.
 35. Bowling MR, Xing D, Kapadia A, Chen YF, Szalai AJ, Oparil S, Hage FG. Estrogen effects on vascular inflammation are age dependent: Role of estrogen receptors. *Arterioscler Thromb Vasc Biol.* 2014; 34:1477-1485.
 36. Aryan L, Younessi D, Zargari M, Banerjee S, Agopian J, Rahman S, Borna R, Ruffenach G, Umar S, Eghbali M. The role of estrogen receptors in cardiovascular disease. *Int J Mol Sci.* 2020; 21.
 37. Vitale C, Miceli M, Rosano GM. Gender-specific characteristics of atherosclerosis in menopausal women: Risk factors, clinical course and strategies for prevention. *Climacteric.* 2007; 10 Suppl 2:16-20.
 38. Qin Y, Santos HO, Khani V, Tan SC, Zhi Y. Effects of dehydroepiandrosterone (DHEA) supplementation on the lipid profile: A systematic review and dose-response meta-analysis of randomized controlled trials. *Nutr Metab Cardiovas.* 2020; 30:1465-1475.
 39. Anagnostis P, Stevenson JC, Crook D, Johnston DG, Godsland IF. Effects of menopause, gender and age on lipids and high-density lipoprotein cholesterol subfractions. *Maturitas.* 2015; 81:62-68.
 40. Goh VH, Tong TY, Mok HP, Said B. Differential impact of aging and gender on lipid and lipoprotein profiles in a cohort of healthy Chinese Singaporeans. *Asian J Androl.* 2007; 9:787-794.
 41. Noyan V, Yucel A, Sagsoz N. The association of androgenic sex steroids with serum lipid levels in postmenopausal women. *Acta Obstet Gynecol Scand.* 2004; 83:487-490.
 42. Lasco A, Frisina N, Morabito N, Gaudio A, Morini E, Trifiletti A, Basile G, Nicita-Mauro V, Cucinotta D. Metabolic effects of dehydroepiandrosterone replacement therapy in postmenopausal women. *Eur J Endocrinol.* 2001; 145:457-461.
 43. Rutkowski K, Sowa P, Rutkowska-Talipska J, Kuryliszyn-Moskal A, Rutkowski R. Dehydroepiandrosterone (DHEA): Hypes and hopes. *Drugs.* 2014; 74:1195-1207.
 44. Williams MR, Dawood T, Ling S, Dai A, Lew R, Myles K, Funder JW, Sudhir K, Komesaroff PA. Dehydroepiandrosterone increases endothelial cell proliferation *in vitro* and improves endothelial function *in vivo* by mechanisms independent of androgen and estrogen receptors. *J Clin Endocrinol Metab.* 2004; 89:4708-4715.
 45. Ceconello AL, Trapp M, Hoefel AL, Marques CV, Arbo BD, Osterkamp G, Kucharski LC, Ribeiro MF. Sex-related differences in the effects of high-fat diets on DHEA-treated rats. *Endocrine.* 2015; 48:985-994.
 46. Elraiyah T, Sonbol MB, Wang Z, Khairalseed T, Asi N, Undavalli C, Nabhan M, Altayar O, Prokop L, Montori VM, Murad MH. Clinical review: The benefits and harms of systemic dehydroepiandrosterone (DHEA) in

- postmenopausal women with normal adrenal function: A systematic review and meta-analysis. *J Clin Endocrinol Metab.* 2014; 99:3536-3542.
47. Wang L, Hao Q, Wang YD, Wang WJ, Li DJ. Protective effects of dehydroepiandrosterone on atherosclerosis in ovariectomized rabbits *via* alleviating inflammatory injury in endothelial cells. *Atherosclerosis.* 2011; 214:47-57.
 48. Jankowski CM, Gozansky WS, Van Pelt RE, Wolfe P, Schwartz RS, Kohrt WM. Oral dehydroepiandrosterone replacement in older adults: Effects on central adiposity, glucose metabolism and blood lipids. *Clin Endocrinol (Oxf).* 2011; 75:456-463.
 49. Li L, Ge C, Wang D, Yu L, Zhao J, Ma H. Dehydroepiandrosterone reduces accumulation of lipid droplets in primary chicken hepatocytes by biotransformation mediated *via* the cAMP/PKA-ERK1/2 signaling pathway. *Biochim Biophys Acta Mol Cell Biol Lipids.* 2018; 1863:625-638.
 50. Fujioka K, Kajita K, Wu Z, Hanamoto T, Ikeda T, Mori I, Okada H, Yamauchi M, Uno Y, Morita H, Nagano I, Takahashi Y, Ishizuka T. Dehydroepiandrosterone reduces preadipocyte proliferation *via* androgen receptor. *Am J Physiol Endocrinol Metab.* 2012; 302:E694-704.
 51. Karbowska J, Kochan Z. Fat-reducing effects of dehydroepiandrosterone involve upregulation of ATGL and HSL expression, and stimulation of lipolysis in adipose tissue. *Steroids.* 2012; 77:1359-1365.
 52. Moreau KL, Hildreth KL, Klawitter J, Blatchford P, Kohrt WM. Decline in endothelial function across the menopause transition in healthy women is related to decreased estradiol and increased oxidative stress. *Geroscience.* 2020; 42:1699-1714.
 53. Messner B, Bernhard D. Smoking and cardiovascular disease: Mechanisms of endothelial dysfunction and early atherogenesis. *Arterioscler Thromb Vasc Biol.* 2014; 34:509-515.
 54. Xu S, Yin M, Koroleva M, Mastrangelo MA, Zhang W, Bai P, Little PJ, Jin ZG. SIRT6 protects against endothelial dysfunction and atherosclerosis in mice. *Aging (Albany NY).* 2016; 8:1064-1082.
 55. Savineau JP, Marthan R, Dumas de la Roque E. Role of DHEA in cardiovascular diseases. *Biochem Pharmacol.* 2013; 85:718-726.
 56. Nheu L, Nazareth L, Xu GY, Xiao FY, Luo RZ, Komesaroff P, Ling S. Physiological effects of androgens on human vascular endothelial and smooth muscle cells in culture. *Steroids.* 2011; 76:1590-1596.
 57. Sanchez-Rodriguez MA, Zacarias-Flores M, Arronte-Rosales A, Correa-Munoz E, Mendoza-Nunez VM. Menopause as risk factor for oxidative stress. *Menopause.* 2012; 19:361-367.
 58. Ogunro PS, Bolarinde AA, Owa OO, Salawu AA, Oshodi AA. Antioxidant status and reproductive hormones in women during reproductive, perimenopausal and postmenopausal phase of life. *Afr J Med Arf Sci.* 2014; 43:49-57.
 59. Kolesnikova L, Semenova N, Madaeva I, Suturina L, Solodova E, Grebenkina L, Darenskaya M. Antioxidant status in peri- and postmenopausal women. *Maturitas.* 2015; 81:83-87.
 60. Hildreth KL, Kohrt WM, Moreau KL. Oxidative stress contributes to large elastic arterial stiffening across the stages of the menopausal transition. *Menopause.* 2014; 21:624-632.
 61. Sukhovshin RA, Yepuri G, Ghebremariam YT. Endothelium-derived nitric oxide as an antiatherogenic mechanism: Implications for therapy. *Methodist Debaque Cardiovasc J.* 2015; 11:166-171.
 62. Maiolino G, Rossitto G, Caielli P, Bisogni V, Rossi GP, Calo LA. The role of oxidized low-density lipoproteins in atherosclerosis: The myths and the facts. *Mediators Inflamm.* 2013; 2013:714653.
 63. Torres N, Guevara-Cruz M, Velazquez-Villegas LA, Tovar AR. Nutrition and atherosclerosis. *Arch Med Res.* 2015; 46:408-426.
 64. Brown DI, Griendling KK. Regulation of signal transduction by reactive oxygen species in the cardiovascular system. *Circ Res.* 2015; 116:531-549.
 65. He F, Zuo L. Redox roles of reactive oxygen species in cardiovascular diseases. *Int J Mol Sci.* 2015; 16:27770-27780.
 66. Cheng HH, Hu XJ, Ruan QR. Dehydroepiandrosterone anti-atherogenesis effect is not *via* its conversion to estrogen. *Acta Pharmacol Sin.* 2009; 30:42-53.
 67. Lopez-Marure R, Huesca-Gomez C, Ibarra-Sanchez Mde J, Zentella A, Perez-Mendez O. Dehydroepiandrosterone delays LDL oxidation *in vitro* and attenuates several oxLDL-induced inflammatory responses in endothelial cells. *Inflamm Allergy Drug Targets.* 2007; 6:174-182.
 68. Miyazaki H, Takitani K, Koh M, Inoue A, Tamai H. Dehydroepiandrosterone alters vitamin E status and prevents lipid peroxidation in vitamin E-deficient rats. *J Clin Biochem Nutr.* 2016; 58:223-231.
 69. Curatola AM, Huang K, Naftolin F. Dehydroepiandrosterone (DHEA) inhibition of monocyte binding by vascular endothelium is associated with sialylation of neural cell adhesion molecule. *Reprod Sci.* 2012; 19:86-91.
 70. Yin FJ, Kang J, Han NN, Ma HT. Effect of dehydroepiandrosterone treatment on hormone levels and antioxidant parameters in aged rats. *Genet Mol Res.* 2015; 14:11300-11311.
 71. Kiersztan A, Trojan N, Tempes A, Nalepa P, Sitek J, Winiarska K, Usarek M. DHEA supplementation to dexamethasone-treated rabbits alleviates oxidative stress in kidney-cortex and attenuates albuminuria. *J Steroid Biochem Mol Biol.* 2017; 174:17-26.
 72. Camporez JP, Akamine EH, Davel AP, Franci CR, Rossoni LV, Carvalho CR. Dehydroepiandrosterone protects against oxidative stress-induced endothelial dysfunction in ovariectomized rats. *J Physiol.* 2011; 589:2585-2596.
 73. Kang J, Ge C, Yu L, Li L, Ma H. Long-term administration of dehydroepiandrosterone accelerates glucose catabolism *via* activation of PI3K/Akt-PFK-2 signaling pathway in rats fed a high-fat diet. *PLoS One.* 2016; 11:e0159077.
 74. Taleb-Belkadi O, Chaib H, Zemour L, Fatah A, Chafi B, Mekki K. Lipid profile, inflammation, and oxidative status in peri- and postmenopausal women. *Gynecol Endocrinol.* 2016; 32:982-985.
 75. Jiang F, Zhang X, Lu YM, Li YG, Zhou X, Wang YS. Elevated level of miR-17 along with decreased levels of TIMP-1 and IL-6 in plasma associated with the risk of instant restenosis. *Biosci Trends.* 2019; 13:423-429.
 76. Novella S, Heras M, Hermenegildo C, Dantas AP. Effects of estrogen on vascular inflammation: A matter of timing. *Arterioscler Thromb Vasc Biol.* 2012; 32:2035-2042.
 77. Davis SR, Castelo-Branco C, Chedraui P, Lumsden MA, Nappi RE, Shah D, Villaseca P, Menopause IMSW.

- Understanding weight gain at menopause. *Climacteric*. 2012; 15:419-429.
78. Kadry RW, Adil MS, Newsome AS, Somanath PR. Cisatracurium attenuates LPS-induced modulation of MMP3 and junctional protein expression in human microvascular endothelial cells. *Biosci Trends*. 2021; 15:50-54.
 79. Gutierrez G, Mendoza C, Zapata E, Montiel A, Reyes E, Montano LF, Lopez-Marure R. Dehydroepiandrosterone inhibits the TNF- α -induced inflammatory response in human umbilical vein endothelial cells. *Atherosclerosis*. 2007; 190:90-99.
 80. Ziogas A, Maekawa T, Wiessner JR, Le TT, Sprott D, Troullinaki M, Neuwirth A, Anastasopoulou V, Grossklaus S, Chung KJ, Sperandio M, Chavakis T, Hajishengallis G, Alexaki VI. DHEA inhibits leukocyte recruitment through the regulation of the integrin antagonist DEL-1. *J Immunol*. 2020; 204:1214-1224.
 81. Radziwon-Balicka A, Lesyk G, Back V, *et al.* Differential eNOS-signalling by platelet subpopulations regulates adhesion and aggregation. *Cardiovasc Res*. 2017; 113:1719-1731.
 82. Ramezani Tehrani F, Behboudi-Gandevani S, Ghasemi A, Azizi F. Association between serum concentrations of nitric oxide and transition to menopause. *Acta Obstet Gynecol Scand*. 2015; 94:708-714.
 83. Tehrani FR, Behboudi-Gandevani S, Ghasemi A, Azizi F. Menopause status as the main factor explaining the gender differences of serum nitric oxide concentrations in middle-aged population. *Arch Gynecol Obstet*. 2015; 291:159-163.
 84. Mury WV, Brunini TM, Abrantes DC, Mendes IK, Campos MB, Mendes-Ribeiro AC, Matsuura C. Hyperaggregability and impaired nitric oxide production in platelets from postmenopausal women. *Maturitas*. 2015; 80:75-81.
 85. Simoncini T, Mannella P, Fornari L, Varone G, Caruso A, Genazzani AR. Dehydroepiandrosterone modulates endothelial nitric oxide synthesis *via* direct genomic and nongenomic mechanisms. *Endocrinology*. 2003; 144:3449-3455.
 86. Munoz YC, Gomez GI, Moreno M, Solis CL, Valladares LE, Velarde V. Dehydroepiandrosterone prevents the aggregation of platelets obtained from postmenopausal women with type 2 diabetes mellitus through the activation of the PKC/eNOS/NO pathway. *Horm Metab Res*. 2012; 44:625-631.
 87. Xu X, Wang B, Ren C, Hu J, Greenberg DA, Chen T, Xie L, Jin K. Age-related impairment of vascular structure and functions. *Aging Dis*. 2017; 8:590-610.
 88. Ross MD, Malone E, Florida-James G. Vascular ageing and exercise: Focus on cellular reparative processes. *Oxid Med Cell Longev*. 2016; 2016:3583956.
 89. Liu D, Si H, Reynolds KA, Zhen W, Jia Z, Dillon JS. Dehydroepiandrosterone protects vascular endothelial cells against apoptosis through a Galphai protein-dependent activation of phosphatidylinositol 3-kinase/Akt and regulation of antiapoptotic Bcl-2 expression. *Endocrinology*. 2007; 148:3068-3076.
 90. Leopold JA, Loscalzo J. Cyclic strain modulates resistance to oxidant stress by increasing G6PDH expression in smooth muscle cells. *Am J Physiol Heart Circ Physiol*. 2000; 279:H2477-2485.
 91. Lee CH, Su SC, Chiang CF, Chien CY, Hsu CC, Yu TY, Huang SM, Shieh YS, Kao HW, Tsai CS, Hung YJ, Lin CY. Estrogen modulates vascular smooth muscle cell function through downregulation of SIRT1. *Oncotarget*. 2017; 8:110039-110051.
 92. Thompson AM, Wagner R, Rzczidlo EM. Age-related loss of SirT1 expression results in dysregulated human vascular smooth muscle cell function. *Am J Physiol Heart Circ Physiol*. 2014; 307:H533-541.
 93. Li L, Zhang HN, Chen HZ, *et al.* SIRT1 acts as a modulator of neointima formation following vascular injury in mice. *Circulation Research*. 2011; 108:1180-U1195.
 94. Mountain DJH, Kirkpatrick SS, Cassada DC, Stevens SL, Freeman MB, Goldman MH, Grandas OH. Estrogen and progesterone induce migration, invasion, and proliferation of vascular smooth muscle cells *via* matrix metalloproteinase regulation. In: 2009 First Annual ORNL Biomedical Science & Engineering Conference: Exploring the intersections of interdisciplinary biomedical research (Evans BM, ed.). Oak Ridge, Tennessee, USA, 2009; pp. 132-135.
 95. Chen J, Xu L, Huang C. DHEA inhibits vascular remodeling following arterial injury: A possible role in suppression of inflammation and oxidative stress derived from vascular smooth muscle cells. *Mol Cell Biochem*. 2014; 388:75-84.
 96. Altman R, Motton DD, Kota RS, Rutledge JC. Inhibition of vascular inflammation by dehydroepiandrosterone sulfate in human aortic endothelial cells: Roles of PPAR α and NF- κ B. *Vasc Pharmacol*. 2008; 48:76-84.
 97. Li Y, Yan L, Zhang W, Hu N, Chen W, Wang H, Kang M, Ou H. Suppression of endothelial nitric oxide synthase expression and endothelial cell proliferation by an intronic 27-ntmiRNA and it's a novel link to AP-1. *Am J Transl Res*. 2015; 7:285-297.
 98. Galvagni F, Orlandini M, Oliviero S. Role of the AP-1 transcription factor FOSL1 in endothelial cells adhesion and migration. *Cell Adh Migr*. 2013; 7:408-411.
 99. Tousoulis D, Kampoli AM, Tentolouris C, Papageorgiou N, Stefanadis C. The role of nitric oxide on endothelial function. *Curr Vasc Pharmacol*. 2012; 10:4-18.
 100. Urata Y, Goto S, Kawakatsu M, Yodoi J, Eto M, Akishita M, Kondo T. DHEA attenuates PDGF-induced phenotypic proliferation of vascular smooth muscle A7r5 cells through redox regulation. *Biochem Biophys Res Commun*. 2010; 396:489-494.
 101. Ii M, Hoshiga M, Negoro N, Fukui R, Nakakoji T, Kohbayashi E, Shibata N, Furutama D, Ishihara T, Hanafusa T, Losordo DW, Ohsawa N. Adrenal androgen dehydroepiandrosterone sulfate inhibits vascular remodeling following arterial injury. *Atherosclerosis*. 2009; 206:77-85.
 102. Baulieu EE. Dehydroepiandrosterone (DHEA): A fountain of youth? *J Clin Endocrinol Metab*. 1996; 81:3147-3151.
 103. Mannella P, Simoncini T, Caretto M, Genazzani AR. Dehydroepiandrosterone and cardiovascular disease. *Vitam Horm*. 2018; 108:333-353.
 104. Casson PR, Santoro N, Elkind-Hirsch K, Carson SA, Hornsby PJ, Abraham G, Buster JE. Postmenopausal dehydroepiandrosterone administration increases free insulin-like growth factor-I and decreases high-density lipoprotein: A six-month trial. *Fertil Steril*. 1998; 70:107-110.
 105. Barnhart KT, Freeman E, Grisso JA, Rader DJ, Sammel M, Kapoor S, Nestler JE. The effect of

- dehydroepiandrosterone supplementation to symptomatic perimenopausal women on serum endocrine profiles, lipid parameters, and health-related quality of life. *J Clin Endocrinol Metab.* 1999; 84:3896-3902.
106. Elraiyah T, Sonbol MB, Wang Z, Khairalseed T, Asi N, Undavalli C, Nabhan M, Altayar O, Prokop L, Montori VM, Murad MH. The benefits and harms of systemic dehydroepiandrosterone (DHEA) in postmenopausal women with normal adrenal function: A systematic review and meta-analysis. *J Clin Endocr Metab.* 2014; 99:3536-3542.
 107. Panjari M, Bell RJ, Jane F, Adams J, Morrow C, Davis SR. The safety of 52 weeks of oral DHEA therapy for postmenopausal women. *Maturitas.* 2009; 63:240-245.
 108. Nair KS, Rizza RA, O'Brien P, *et al.* DHEA in elderly women and DHEA or testosterone in elderly men. *N Engl J Med.* 2006; 355:1647-1659.
 109. Gebre-Medhin G, Husebye ES, Mallmin H, Helstrom L, Berne C, Karlsson FA, Kampe O. Oral dehydroepiandrosterone (DHEA) replacement therapy in women with Addison's disease. *Clin Endocrinol (Oxf).* 2000; 52:775-780.
 110. Wu TT, Gao Y, Zheng YY, Ma YT, Xie X. Association of endogenous DHEA/DHEAS with coronary heart disease: A systematic review and meta-analysis. *Clin Exp Pharmacol Physiol.* 2019; 46:984-994.
 111. Wu TT, Chen Y, Zhou Y, Adi D, Zheng YY, Liu F, Ma YT, Xie X. Prognostic value of dehydroepiandrosterone sulfate for patients with cardiovascular disease: A systematic review and meta-analysis. *J Am Heart Assoc.* 2017; 6:e004896.
 112. Eden JA. DHEA replacement for postmenopausal women: Placebo or panacea? *Climacteric.* 2015; 18:439-440.
 113. Davis SR, Panjari M, Stanczyk FZ. DHEA replacement for postmenopausal women. *J Clin Endocr Metab.* 2011; 96:1642-1653.
 114. Genazzani AR, Pluchino N. DHEA replacement for postmenopausal women: Have we been looking in the right direction? *Climacteric.* 2015; 18:669-671.
 115. Dhatariya KK, Nair KS. Dehydroepiandrosterone: Is there a role for replacement? *Mayo Clin Proc.* 2003; 78:1257-1273.
 116. Friis Berntsen C, Rootwelt P, Dahm AEA. Bias in animal studies of estrogen effects on cardiovascular disease: A systematic review and meta-analysis. *Res Pract Thromb Haemost.* 2021; 5:e12507.
 117. Genazzani AR, Pluchino N. DHEA therapy in postmenopausal women: The need to move forward beyond the lack of evidence. *Climacteric.* 2010; 13:314-316.
 118. Scheffers CS, Armstrong S, Cantineau AE, Farquhar C, Jordan V. Dehydroepiandrosterone for women in the peri- or postmenopausal phase. *Cochrane Database Syst Rev.* 2015; 1:CD011066.
 119. Marsden J, British Menopause S. British Menopause Society consensus statement: The risks and benefits of HRT before and after a breast cancer diagnosis. *Post Reprod Health.* 2019; 25:33-37.
 120. Pluchino N, Carmignani A, Cubeddu A, Santoro A, Cela V, Alcalá TE. Androgen therapy in women: For whom and when. *Archives of Gynecology and Obstetrics.* 2013; 288:731-737.
 121. Hodis HN, Mack WJ, Henderson VW, Shoupe D, Budoff MJ, Hwang-Levine J, Li Y, Feng M, Dustin L, Kono N, Stanczyk FZ, Selzer RH, Azen SP. Vascular effects of early versus late postmenopausal treatment with estradiol. *N Engl J Med.* 2016; 374:1221-1231.
 122. Weiss EP, Villareal DT, Fontana L, Han DH, Holloszy JO. Dehydroepiandrosterone (DHEA) replacement decreases insulin resistance and lowers inflammatory cytokines in aging humans. *Aging (Albany NY).* 2011; 3:533-542.
- Received August 2, 2021; Revised October 28, 2021; Accepted November 5, 2021.
- §These authors contributed equally to this work.
- *Address correspondence to:
Ling Wang, Laboratory for Reproductive Immunology, Obstetrics and Gynecology Hospital of Fudan University, 419 Fangxie Road, Shanghai 200011, China.
E-mail: Dr.wangling@fudan.edu.cn
- Released online in J-STAGE as advance publication November 10, 2021.